

(lit.<sup>26</sup> m.p. 181–182°); infrared spectrum (KBr), OH 2.85–(m)  $\mu$  and C=O 5.75(s)  $\mu$ .

Acidification of the combined base washings, extraction with ether, drying, and solvent evaporation led to 50 mg. of non-crystalline acidic material,  $[\alpha]_D -38^\circ$  (EtOH); ultraviolet spectrum (95% ethanol),  $\lambda_{\max}$  241 m $\mu$ . The acidic substance was dissolved in 0.1 ml. of acetone, 2 drops of di-*n*-amylamine (b.p. 190–192°) added<sup>27</sup> and the solution cooled. Filtration of the crystalline precipitate and crystallization from acetone furnished a salt,  $[\alpha]_D -53^\circ$  (EtOH), whose infrared spectrum was identical with that of a

(37) Cf. G. C. Harris and T. F. Sanderson, *THIS JOURNAL*, **70**, 334 (1948).

freshly prepared sample of the di-*n*-amylamine salt of abietic acid,  $[\alpha]_D -59^\circ$  (EtOH).

Identical acid treatment and work-up of 100 mg. of isopimaric acid yielded 17 mg. of a non-crystalline mixture of 5- and 6-lactones, 26 mg. of hydroxylactone, m.p. 160–175°, increased to 180–181° after crystallization from petroleum ether–acetone, no depression on admixture with above hydroxylactone, identical infrared spectra, and 45 mg. of acid,  $[\alpha]_D -45^\circ$  (EtOH), ultraviolet spectrum (95% ethanol),  $\lambda_{\max}$  241 m $\mu$ , whose di-*n*-amylamine salt,  $[\alpha]_D -58^\circ$  (EtOH), had an infrared spectrum identical with that of the abietic acid salt.

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## Aminodihydrofuramides from 3-Amino-1-propynes and Carbon Monoxide

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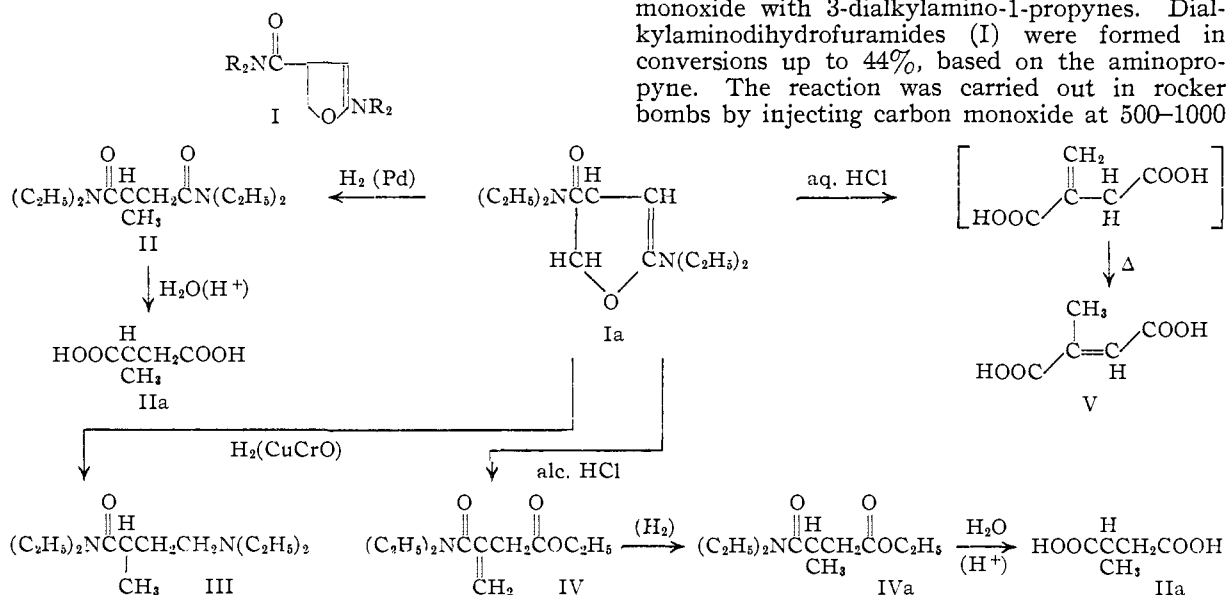
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The reaction of 3-diethylamino-1-propyne with carbon monoxide has given *N,N*-diethyl-5-diethylamino-2,3-dihydro-3-furamide in 44% yield. The synthesis was carried out by injecting carbon monoxide at 500–1000 atm. into the aminopropyne dissolved in a ketone solvent at a temperature of 125°. Catalytic amounts of dicobalt octacarbonyl were necessary for the synthesis. The reaction has also been extended to several other aminopropynes. The products undergo facile ring cleavage with hydrogen, hydrogen chloride or water to give succinic acid derivatives. The mechanism for this unusual transformation is unknown, in part because other products of the reaction were isolable only as intractable residues.

The literature contains many references to the reactions of acetylene or substituted acetylenes with carbon monoxide. These reactions generally involved various metallic carbonyls as catalysts or

carbon monoxide needed for the synthesis, several 3-dialkylamino-1-propynes were converted into the 2,5-bis-(dialkylaminomethyl)-hydroquinones.<sup>2</sup>

This paper describes a new reaction of carbon monoxide with 3-dialkylamino-1-propynes. Dialkylaminodihydrofuramides (I) were formed in conversions up to 44%, based on the aminopropyne. The reaction was carried out in rocker bombs by injecting carbon monoxide at 500–1000



reactants, and the products were mainly acrylic compounds or hydroquinones.<sup>1</sup> Although a number of functionally substituted acetylenes have been studied in these reactions, only one reference to the interaction of 3-dialkylamino-1-propynes with carbon monoxide appears to have been reported. With iron carbonyl hydride furnishing the

atm. into a solution of the aminopropyne at 125°. Ketones such as acetone or cyclohexanone were the best solvents tested. Catalytic amounts of dicobalt octacarbonyl were necessary for the synthesis.

The novel products were identified mainly on the basis of the chemical evidence indicated schematically below. *N,N*-Diethyl-5-diethylamino-2,3-dihydro-3-furamide (Ia) undergoes facile ring cleavage at the bond in the 1,2-position. Open-chain compounds, all of which may be considered to be derivable from methylsuccinic acid, were formed in

(1) (a) J. W. Copenhaver and M. H. Bigelow, "Acetylene and Carbon Monoxide Chemistry," Reinhold Publishing Corp., New York, N. Y., 1949; (b) J. W. Reppe, *et al.*, *Ann.*, **582**, 1 (1953); (c) E. R. H. Jones, T. Y. Shen and M. C. Whiting, *J. Chem. Soc.*, 230 (1950); 48, 763, 766 (1951); (d) E. R. H. Jones, G. H. Whitham and M. C. Whiting, *ibid.*, 1865 (1954).

(2) Reference 1a, p. 293; J. W. Reppe, *et al.*, *Ann.*, **582**, 142 (1953).





tions of ether. The ether extracts and the top layer were then combined and dried over potassium carbonate. Ether was removed on a steam-bath, and 126 g. of 3-diethylamino-1-propyne, distilling at 119–120°, was obtained (76% yield,  $n_D^{25}$  1.4288). This procedure has obvious advantages over the literature method which requires diethylamine acetate.<sup>7</sup>

**3-(4-Morpholino)-1-propyne.**—3-(4-Morpholino)-1-propyne, b.p. 89–92° (38 mm.)  $n_D^{25}$  1.4723, was obtained in 66% yield by utilizing the procedure described above. This propyne was identified by infrared and nuclear magnetic resonance spectra and analytical data. *Anal.* Calcd. for  $C_7H_{11}NO$ : C, 67.2; H, 8.9; N, 11.2; mol. wt., 125. Found: C, 67.7; H, 8.9; N, 10.9; mol. wt., 123, 128. The infrared spectrum indicated  $\equiv CH$  (3.0  $\mu$ ), saturated CH (3.4, 3.5 and 3.7  $\mu$ ), and  $-C\equiv C-$  (4.75  $\mu$ ). The nuclear magnetic resonance spectrum indicated acetylenic hydrogen and two types of methylene hydrogen.

**3-Diethylamino-3-methyl-1-propyne**, b.p. 126–128°,  $n_D^{25}$  1.4273, was prepared in 33% yield from acetylene and diethylamine by a published procedure.<sup>8</sup>

(7) Ref. 1a, p. 110; also, Reppe, *Ann.*, **596**, 1 (1955).

(8) C. Gardner, V. Kerrigan, J. D. Rose and B. C. L. Weedon, *J. Chem. Soc.*, 780 (1949).

**Reaction of 3-Diethylamino-1-propyne with Dicobalt Octacarbonyl.**—3-Diethylamino-1-propyne (0.19 g.) in acetone (25 ml.) was added to dicobalt octacarbonyl (0.57 g.) in the standard Orsat apparatus for measuring gases. There was collected 63.3 ml. of carbon monoxide, making the necessary corrections for acetone vapor (87% of theory).

Commercial-grade acetylene was purified according to a previously described procedure.<sup>9</sup> The infrared spectra were determined on a Perkin-Elmer 21 double-beam spectrometer. The ultraviolet spectra were determined on a Cary model 11 spectrophotometer. The proton magnetic resonance spectra were obtained using a Varian high-resolution n.m.r. spectrometer and electromagnet at frequencies of 40 Mc. and fields of 10,000 gauss, respectively. The spectra were calibrated in terms of displacements in cycles per second (c.p.s.) from the proton resonance of water. Positive values are on the low field side of water, and negative values are on the high field side. Calibration was accomplished by superimposing an audiofrequency on the sweep field to produce side band peaks to the water resonance. Yields of the dihydrofuramides were calculated on the basis of the amount of dialkylamino-propyne consumed.

(9) J. C. Sauer, *THIS JOURNAL*, **79**, 5314 (1957).

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[CONTRIBUTION FROM THE ROCKEFELLER INSTITUTE FOR MEDICAL RESEARCH]

## Characterization Studies with Subtilin

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Further purification studies of subtilin A by countercurrent distribution (c.c.d.) are reported. Molecular weight determination by the method of partial substitution has conclusively shown that subtilin A approximates 3300 in molecular weight. This and quantitative amino acid analyses are consistent with an amino acid formula of Asp, Pro, Gly, Ala, Val, Ileu, Leu, Phe, Lys, Lan,  $\beta$ -MeLan, Glu, Try, Sar, Sarcosine, not heretofore reported, has been shown to be an N-terminal group. Subtilin A is a pentacyclic peptide with a side chain.

A number of different polypeptide antibiotics<sup>2</sup> in the molecular weight range up to 1500 have now been well characterized chemically and reasonably certain cyclic structures have been proposed for them. Although larger ones are known they have not thus far been studied as carefully. Subtilin<sup>3</sup> produced by a particular strain of *Bacillus subtilis* is the best characterized and perhaps the most readily available member of the larger size group.

A preliminary survey of the excellent chemical work already done with subtilin<sup>4–9</sup> indicated that it probably was sufficiently well characterized for further structural study. Nonetheless, because of the effort required in such an undertaking it seemed wise to repeat part of the work using different methods. This paper will report studies of this nature and some new observations bearing on the structure of the peptide.

### Experimental

**Materials.**—Three samples of subtilin all received from the Western Regional Laboratory have been studied thus far. One was a 5-g. sample (Lot 317) received from Dr. Harold Olcott in 1950. Two more samples were received

from Dr. Alderton in 1957. One of these, 152F, was similar to 317 but freshly prepared whereas 317 had been stored for about 10 years. The other was relatively pure subtilin A obtained by silica gel partition chromatography of 152F. Dr. Alderton had found it to give a single band by countercurrent distribution at 180 transfers (system = 20% acetic acid, 5, *n*-butanol, 4) with close agreement to a calculated curve. It behaved the same way in our hands.

**Fractionation Studies.**—Fractionation was accomplished by countercurrent distribution in a 1000 tube (2 ml. lower phase) automatic train of the type previously described<sup>10</sup> in the Alderton system. The charge was 1 g. of sample 152F initially scattered in the first twenty tubes of the train. Each transfer required 2.5 minutes including 5 strokes for equilibration and 1.5 minutes for separation of the phases. 5 strokes seemed to be sufficient for equilibration. The temperature of the train was 25°.

After 1000 transfers the upper patterns of Fig. 1 were obtained by optical density measurement at 288  $m\mu$  of the upper and lower phases in a Beckman quartz spectrophotometer. Several small bands as indicated and a larger one on the right, C<sub>3</sub>, were removed from the train. After these tubes were filled with fresh phases the distribution was continued to 2540 transfers by the recycling procedure.

Analysis by optical density now gave the lower patterns of Fig. 1. A plot of the partition ratio across the main band is given above the distribution curve.

Cuts labeled A<sub>3</sub> and B<sub>3</sub> were taken as indicated on the chart. They were concentrated in a rotary evaporator until all the butanol phase was removed and lyophilized from the aqueous solution. The preparation from cut A<sub>3</sub> was the material used for the characterization studies reported in this paper. It corresponds to subtilin A.

*Anal.* The sample (from the run of Fig. 1) lost 18.2% on drying at 100° *in vacuo*. Found: C, 52.97; H, 6.99; N, 16.06; S, 4.76; N-CH<sub>3</sub>, 1.92.

- (1) Fellow of the National Foundation for Infantile Paralysis.
- (2) L. C. Craig, *Proc. 3rd Int. Cong. Biochem.*, Brussels, 1955.
- (3) K. P. Dimick, G. Alderton, J. C. Lewis, H. D. Lightbody and H. L. Fevold, *Arch. Biochem.*, **15**, 1 (1947).
- (4) J. C. Lewis and N. S. Snell, *THIS JOURNAL*, **73**, 4812 (1951).
- (5) N. G. Brink, J. Mayfield and K. Folkers, *ibid.*, **73**, 330 (1951).
- (6) G. Alderton and H. L. Fevold, *ibid.*, **73**, 463 (1951).
- (7) J. F. Carson, *ibid.*, **74**, 1480 (1952).
- (8) G. Alderton, *ibid.*, **75**, 2391 (1953).
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- (10) L. C. Craig, W. Hausmann, E. H. Ahrens, Jr., and E. P. Harfenist, *Anal. Chem.*, **23**, 1236 (1951).